The Molecular Basis of Cystathionine β-Synthase Deficiency in Dutch Patients with Homocystinuria: Effect of CBS Genotype on Biochemical and Clinical Phenotype and on Response to Treatment

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Summary

Homocystinuria due to cystathionine β -synthase (CBS) **deficiency, inherited as an autosomal recessive trait, is the most prevalent inborn error of methionine metabolism. Its diverse clinical expression may include ectopia lentis, skeletal abnormalities, mental retardation, and premature arteriosclerosis and thrombosis. This variability is likely caused by considerable genetic heterogeneity. We investigated the molecular basis of CBS deficiency in 29 Dutch patients from 21 unrelated pedigrees and studied the possibility of a genotype-phenotype relationship with regard to biochemical and clinical expression and response to homocysteine-lowering treatment. Clinical symptoms and biochemical parameters were recorded at diagnosis and during long-term followup. Of 10 different mutations detected in the CBS gene,** 833T \rightarrow C (I278T) was predominant, present in 23 (55%) **of 42 independent alleles. At diagnosis, homozygotes for** this mutation $(n = 12)$ tended to have higher homocy**steine levels than those seen in patients with other gen**otypes ($n = 17$), but similar clinical manifestations. Dur**ing follow-up, I278T homozygotes responded more efficiently to homocysteine-lowering treatment. After 378 patient-years of treatment, only 2 vascular events were recorded; without treatment, at least 30 would** have been expected $(P < .01)$. This intervention in Dutch **patients significantly reduces the risk of cardiovascular disease and other sequelae of classical homocystinuria syndrome.**

Introduction

Homocystinuria due to cystathionine β -synthase (CBS; L-serine hydrolyase [adding homocysteine]) deficiency (MIM 236200) is the most common inborn error in methionine metabolism. CBS is a pyridoxal 5 -phosphate (PLP)–dependent enzyme and condenses homocysteine and serine to cystathionine, an irreversible step in transsulfuration (Mudd et al. 1995).

Carson and Neill (1962) first described homocystinuria in mentally retarded individuals in Northern Ireland. Soon thereafter, it was shown that the primary defect in homocystinuria was an enzymatic defect of CBS (Mudd et al. 1964), with a recessive mode of inheritance (Finkelstein et al. 1964). The clinical manifestation of CBS deficiency is diverse, and four major organ systems are predominantly involved: the eye (high myopia and ectopia lentis), the skeleton (osteoporosis, scoliosis, and Marfanoid features), the vascular system (premature arteriosclerosis and thromboembolism), and the CNS (mental retardation, convulsions, and psychiatric disturbances).

Biochemically, patients with CBS deficiency are characterized by severe hyperhomocysteinemia and homocystinuria, hypermethioninemia, and decreased plasma cysteine levels. Furthermore, the CBS activities measured in either liver biopsy specimens (Mudd et al. 1964), cultured fibroblasts (Uhlendorf and Mudd 1968), or phytohemagglutinin-stimulated lymphocytes (Goldstein et al. 1972) are mostly well below the range of CBS activities observed in controls and heterozygotes for CBS deficiency. The first choice of therapy in CBS-deficient patients consists of administration of supraphysiological doses of pyridoxine (vitamin B_6), the precursor of PLP, the cofactor of CBS. A large international survey of >600 patients with homocystinuria showed that ∼50% of the patients responded to high doses of pyridoxine with a substantial reduction in blood homocysteine concentrations (Mudd et al. 1985). Pyridoxine-nonresponsive patients usually are more severely affected than pyridoxine-

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Table 1

CBS Mutations in Dutch Homocystinurics

1111G→A V371M + *Nla*III 2/42 1301C \rightarrow A T434N None 1/42
1330G \rightarrow A D444N $-TaqI$ 2/42

 $1471C \rightarrow T^b$ R491C $+ Bg/I$ 2/42

^a In *independent* alleles.

 $1330G\rightarrow A$

^b Mutations were observed in *cis* in one patient.

*Taq*I 2/42

responsive patients and are concomitantly treated with combinations of folic acid, hydroxycobalamin, and betaine, to stimulate remethylation of homocysteine to methionine.

Data on the clinical efficacy of homocysteine-lowering treatment are scarce. In pyridoxine-responsive patients and early-treated pyridoxine-nonresponsive patients, such treatment has reduced the number of initial thromboembolic events (Mudd et al. 1985). A recent report by Wilcken and Wilcken (1997) showed that treatment that effectively lowered plasma homocysteine concentrations markedly reduced the cardiovascular risk in a group of 32 patients with pyridoxine-responsive and pyridoxine-nonresponsive homocystinuria.

The human CBS gene, which has been mapped to 21q22.3 (Müncke et al. 1988), encodes a CBS subunit of 63 kD, which assembles into a homotetrameric protein (Kraus et al. 1978). So far, >60 mutations have been detected in the CBS gene (Kraus 1998), and functional relevance has been tested for some of them in either a bacterial (de Franchis et al. 1994; Marble et al. 1994; Kluijtmans et al. 1996*a*) or a yeast (Kruger and Cox 1995) expression system. Although most mutations seem to be private or restricted to only a few pedigrees, three mutations are relatively common among patients with homocystinuria. An $833T\rightarrow C$ transition (I278T) (Kozich and Kraus 1992) has been found in alleles from homocystinuric patients of different ethnic backgrounds and has been reported to be associated with pyridoxine responsiveness and a relatively mild clinical phenotype when present in homozygous state (Shih et al. 1995). On the other hand, a 919G \rightarrow A transition (G307S) is related to a more severe clinical phenotype and has been detected mainly in alleles from homocystinuric patients of Celtic origin (Gallagher et al. 1995). A third relatively common CBS mutation, an IVS11-2A \rightarrow C splice mutation, which results in skipping exon 12, has been detected in patients from Central and Eastern Europe (Kraus 1998).

In the present study, we investigated the molecular basis of homocystinuria due to CBS deficiency in 29 Dutch homocystinuria patients from 21 unrelated pedigrees, and we studied a possible relationship between CBS genotype and biochemical and clinical phenotype. We therefore measured homocyst(e)ine concentrations at diagnosis, upon pyridoxine treatment, and upon maximal treatment, with pyridoxine and folic acid, with or without betaine, and then recorded whether homocysteine-lowering treatment had been able to prevent further clinical events or symptoms.

Patients, Material, and Methods

Patients

We studied 29 patients with homocystinuria due to CBS deficiency, from 21 unrelated pedigrees. Patients were initially diagnosed on the basis of clinical manifestations of homozygous CBS deficiency, in combination with a quantitative determination of severe hyperhomocysteinemia and hypermethioninemia. In two patients (19 and 29), the diagnosis was made only on the basis of severe homocystinuria, which was demonstrated by qualitative urine analyses. Homozygous CBS deficiency in these two patients has been confirmed by detection of a homozygous mutation (patient 19) and by the clinical manifestation of ectopia lentis (19 and 29), never observed in heterozygotes for CBS deficiency. Furthermore, the parents of patient 29 showed an abnormal response to a methionine-loading test, comparable to that observed in obligate heterozygotes. Homocysteine-lowering treatment was initiated in all patients immediately after the diagnosis had been made. Each patient was seen on a regular basis (once or twice each year) by two of us (Boers and Cruysberg), and the biochemical efficacy of homocysteine-lowering therapy was determined by measurement of homocyst(e)ine and methionine in blood. At diagnosis and during long-term follow-up, clinical manifestations were recorded by means of routine clinical, radiographic, or scintigraphic examination procedures.

Pyridoxine Responsiveness

Pyridoxine responsiveness was examined after 6 weeks of treatment with vitamin B_6 , 750 mg/day in adults or 200–500 mg/day in children. Patients in whom non–protein-bound serum homocysteine had decreased to $\langle 20 \mu m o l / L$, or total plasma homocysteine (protein*and* non–protein-bound) to $\lt 50 \mu$ mol/L, were classified as pyridoxine responsive. All other patients were categorized as pyridoxine-nonresponsive homocystinurics.

Biochemical Analysis

Homocystine, homocysteine-cysteine–mixed disulfide, and methionine levels in serum were determined as described by Boers et al. (1983). The amount of non–protein-bound homocysteine was calculated as the sum of twice the concentration of homocystine plus the concentration of the homocysteine-cysteine–mixed disulfide moiety. Since 1990, total plasma homocysteine concentrations (i.e., the total amount of protein- *and* non–protein-bound homocysteine moieties) were determined as described by Te Poele-Pothoff et al. (1995).

CBS activities in extracts of cultured fibroblasts were measured as initially described by Fowler et al. (1978), with some modifications (Boers et al. 1985). These CBS activities were measured with and without addition of 1 mM PLP to the incubation mixture.

Mutation Analysis

The procedure for mutation analysis of the CBS gene in patients with homocystinuria has been described elsewhere (Kluijtmans et al. 1996*a*). In brief, genomic DNA was isolated from peripheral blood leukocytes (Miller et al. 1988) and stored at 4°C. Total RNA was extracted from cultured fibroblasts by the method of Chomczynski and Sacchi (1987) and was stored as an ethanol precipitate at -80° C. We used 1–5 μ g total RNA for firststrand cDNA synthesis with Superscript II Reverse Transcriptase (Life Technologies). First-strand cDNA was used as a template in PCR amplification reactions to amplify the CBS-encoding region in multiple overlapping cDNA fragments. These fragments were subsequently sequenced on an ABI 377 automated DNA sequencer (Applied Biosystems) with the Taq Dye Deoxy Terminator Cycle Sequencing Kit. All cDNA fragments were sequenced on both strands, and mutations were confirmed at the genomic DNA level by restriction enzyme analysis or DNA sequencing. Restriction enzymes were purchased from Life Technologies or from New England Biolabs and were used according to the manufacturers' recommendations. Screening for $833T \rightarrow C$ was performed as described elsewhere (Kluijtmans et al. 1996*b*). Using this procedure, we were able to discriminate between the real $833T\rightarrow C$ carriers and those with an 844ins68 duplication variant (Sebastio et al. 1995).

Cloning and Expression of Mutations

cDNA fragments containing the presumed functional mutations were amplified by PCR and subcloned into an expression cartridge as described elsewhere (de Franchis et al. 1994). Recombinant clones were selected and sequenced to verify the integrity of the cloned fragment. CBS expression was induced upon addition of isopropylb-thiogalactopyranoside (Kozich and Kraus 1992). The CBS assay was performed without addition of bovine serum albumin to the incubation mixture, to which cystathionine was added at a final concentration of 2 mM.

Statistics

Differences in clinical manifestation of CBS deficiency and efficacy of homocysteine-lowering treatment between separate groups were assessed by Yates' corrected x^2 test. Differences in age at diagnosis, homocysteine, and methionine concentrations were assessed by nonparametric Wilcoxon–Mann-Whitney U tests. All *P* values reported are two-tailed, and $P < .05$ was considered statistically significant.

Results

Study Group

Twenty-nine homocystinuria patients from 21 pedigrees were included in this study. Six families had two affected siblings; one family had three affected siblings. The male–female ratio was 16:13. The diagnosis of CBS deficiency was established at a mean age of 26 years (median, 23 years [range 4–60]).

Genetic Basis of Homocystinuria

The CBS gene of the patients with homocystinuria was analyzed for mutations, either by direct sequencing of reverse transcription–PCR–amplified fragments (in 14 patients) or by RFLP analysis of genomic DNA fragments, to screen for previously recognized mutations (in 15 patients). The molecular basis of homocystinuria was resolved in 25 (86%) of the 29 patients. Overall, 10 different mutations were found (table 1), of which the $833T\rightarrow C$ mutation was the most prevalent. This mutation was observed in 23 (55%) of 42 independent alleles. In four patients, including one in whom the entire cDNA was sequenced, only one mutation in heterozygous state was found; the mutation in the second allele has yet to be found (table 2).

CBS Activities in Cultured Fibroblasts

CBS activities were measured in extracts of cultured fibroblasts in 12 healthy controls, in 9 CBS-deficient patients homozygous for I278T, and in 12 patients with another CBS genotype (table 2). Without PLP addition to the incubation mixture, the mean $(\pm SD)$ CBS activity in homozygotes was .17 (\pm .37) nmol cystathionine/mg protein/h, <2.5% of the control mean (7.4 [\pm 5.1] nmol cystathionine/mg protein/h). One homocystinuric patient (18) clearly exhibited CBS activities, in the range of obligate heterozygotes, and has been described elsewhere (Kluijtmans et al. 1996*a*).

Table 2

NOTE.—A plus sign (+) indicates pyridoxine responsive, and a minus sign (-) indicates pyridoxine nonresponsive; $ND = not determined$.

Cystathionine β -synthase activity expressed in nmol cystathionine formed/mg protein/h.

 $\frac{1}{2}$ Mean (\pm SD) CBS activity in fibroblasts of healthy controls: 7.4 (\pm 5.1).

The mean CBS activity in fibroblasts of homozygotes for I278T $(n = 9)$ versus patients with other genotypes $(n = 12)$ was 0.05 ± 0.07 versus 0.13 ± 0.18 nmol cystathionine/mg protein/h ($P = .28$), and 0.13 \pm 0.14 versus 0.42 ± 0.38 *nmol cystathionine/mg protein/h (P =* $\frac{1}{2}$.07) in the assay without and with, respectively, 1 mM PLP. Patient 18 was excluded in these calculations.

In Vitro Expression of Mutations

The functional relevance of seven mutations was investigated in an *Escherichia coli* expression system: six constructs contained a single mutation, and one construct contained the 1105C \rightarrow T (R369C) and 1471C \rightarrow T (R491C) mutations in *cis.* All mutated constructs, except the one containing $1330G\rightarrow A$ (D444N) (Kluijtmans et al. 1996*a*), showed a reduction in CBS activity of >90%, demonstrating the detrimental effects of each construct on CBS activity (fig. 1). Two mutations $(373C \rightarrow T)$ [R125W] and 1301C \rightarrow A [T434N]) have not yet been functionally tested.

Pyridoxine Responsiveness

Fourteen (48%) of 29 patients were classified as pyridoxine responsive and 9 (31%) patients as pyridoxine nonresponsive, on the basis of the criteria described in the Patients, Material, and Methods section. In six patients (12, 13, 16, 21, 28, and 29), responsiveness to pyridoxine alone could not be assessed. In view of their extremely high homocysteine levels at diagnosis, these patients were treated directly with a combination of therapeutic regimens (pyridoxine and folic acid, with or without betaine). Pyridoxine responders were diagnosed at a mean age of 29 years (median, 26 years [range, 7–54 years]), and nonresponders at a mean age of 19 years (median, 16 years [range, $4-30$ years]; $P = .08$).

Seven (58%) of 12 homozygotes for the I278T mu-

Figure 1 CBS activities measured in an *E. coli* expression system. Mutations were introduced as described in the Patients, Material, and Methods section. Bacterial lysates were assayed for CBS activity, and the mean CBS activity of a control construct (40.3 nmol cystathionine formed/mg protein/h) has been set to 100%. Each bar represents another mutated construct and is the mean of two independently performed CBS assays.

tation showed in vivo pyridoxine responsiveness, 3 (25%) were nonresponders, and, in 2 (17%) patients, this specific responsiveness could not be assessed. In 17 individuals with other genotype combinations, including compound heterozygotes for I278T, these numbers were 7 (41%), 6 (35%), and 4 (24%), respectively (χ^2 = 0.13; $P = .7$). Conversely, in 14 pyridoxine-responsive patients, 18 (64%) of 28 alleles carried the I278T mutation (7 homozygotes, 4 heterozygotes), versus 6 (33%) I278T alleles in 9 nonresponsive patients (3 homozygotes) ($\chi^2 = 1.39$; $P = .5$). There was an absolute concordance of pyridoxine responsiveness between siblings.

Clinical Characteristics at Diagnosis

The clinical manifestation of homocystinuria due to CBS deficiency in this study group is very diverse and is depicted in detail in table 3. At diagnosis, some patients (2, 13, and 15) showed virtually no clinical symptoms and were investigated because of a homocystinuric sibling. In other patients (6, 19, 22, 23, and 25), all four major organ systems were involved. Ocular abnormalities and skeletal abnormalities were the most consistent findings among these 29 patients with homocystinuria: 25 (86%) suffered from either high myopia or ectopia lentis. Twenty-six (90%) patients exhibited osteopo-

rosis, scoliosis, or Marfanoid features, and 13 (45%) patients had complications in the vascular system. The CNS was involved in 16 (55%) patients, and psychiatric illness was noticed in only 4 (14%) patients. No significant differences in clinical presentation of CBS deficiency were observed between I278T homozygotes and patients with other genotypes (data not shown).

Biochemical Characteristics at Diagnosis

At diagnosis, non–protein-bound homocysteine concentrations had been measured in 21 patients and total homocysteine concentrations in 6 patients. In two patients, no baseline homocysteine blood levels were available; in these two cases the initial diagnosis had been made in 1975, by means of qualitative examination of urine only. The mean non–protein-bound homocysteine concentration was 135 μ mol/L (range, 42–266 μ mol/L; $n = 21$) and mean total plasma homocysteine concentration was 240 μ mol/L (range, 134–299 μ mol/L; $n =$ 6). Mean serum methionine concentration was 130 μ mol/L (range, 52–549 μ mol/L; $n = 23$). Homozygotes for the I278T mutation tended to have a higher mean homocysteine concentration than homocystinuria patients with other genotypes $(160 \pm 73 \mu mol/L)$ versus 116 ± 57 μ mol/L non–protein bound; $P = .16$), whereas methionine concentrations were not significantly different between both genotype groups (89 \pm 35 μ mol/L versus $152 \pm 166 \mu$ mol/L; $P = .49$).

Response to Homocysteine-Lowering Treatment

Long-term homocysteine-lowering therapy (mean term, 13 years [range, 1–29 years]) consisted of maximally 750 mg pyridoxine. Fifteen (56%) patients were concomitantly treated with 5 mg folic acid per day, and eight (30%) patients also with 6 g betaine per day. Only one patient (17) had a methionine-restricted diet, with a methionine content of 600 mg/d. Intramuscular injections with 1 mg hydroxycobalamin every 1–2 mo were given to four patients, because of the development of a vitamin B_{12} deficiency. Two patients could not be followed after diagnosis had been made: patient 6 refused treatment and patient 13 moved to another country. Hence, follow-up was recorded in 27 (93%) of the patients with homocystinuria. The mean length of followup was 11 years (range, $1-20$ years; $n = 11$) in homozygotes for the I278T mutation and 16 years (range, 3–27 years; $n = 16$ in patients with other genotypes $(P = .09)$.

Biochemically, pyridoxine treatment resulted in a marked decrease in homocysteine concentrations of 90% in homozygotes for the I278T mutation and of 67% in patients with other genotypes $(P < .02)$. Extended intervention with folic acid, with or without betaine, further decreased homocysteine concentrations in

Table 3

Clinical Presentation of CBS Deficiency at Diagnosis

Manifested by at least extreme height and arachnodactyly.

 $^{\rm b}$ Peripheral arterial disease in patients 7, 10–12, and 28; cerebrovascular disease in patients 12 and 25; and coronary artery disease in patients 6 and 28.

 ϵ Deep venous thrombosis in patients 1, 7, 19, 23, and 25; pulmonary embolia in patients 9 and 22; and sinus sagittalis thrombosis in patient 21.
d Severe mental retardation in patients 17 and 19; mild mental retardation (not less than 80) in 10 other patients, as indicated.

both genotype groups, by 84% and 47%, respectively $(P < .05)$. Homocysteine concentrations normalized (i.e., non–protein-bound homocysteine $\langle 20 \mu m o/L \rangle$, or total plasma homocysteine $\langle 50 \mu mol/L \rangle$ in 21 (78%) of 27 patients. Methionine concentrations decreased in both genotype groups upon pyridoxine therapy, by 56% and 33%, respectively $(P = .15)$. Additional treatment with folic acid and betaine, to stimulate homocysteine remethylation, left methionine concentrations in homozygotes for the I278T mutation virtually unchanged, whereas methionine levels substantially increased in patients with other genotypes.

During long-term treatment, 9 (82%) of 11 homozygotes for I278T continuously presented non–proteinbound homocysteine concentrations $\langle 20 \mu m o I/L \rangle$ or total homocysteine concentrations $\lt 50 \ \mu$ mol/L, versus only 6 (38%) of 16 patients with another genotype $(P = .06)$. Mean $(\pm SD)$ non–protein-bound homocysteine during long-term follow-up was 13 ± 13 μ mol/L in I278T homozygotes versus 23 ± 18 µmol/L in patients with other genotypes ($P = .31$). For total homocysteine levels, these concentrations were 30 ± 16 µmol/L and 67 ± 16 38 μ mol/L, respectively ($P < .01$). Despite extended therapy, four patients (14, 15, 17, and 18), all without the I278T mutation, persistently had total plasma homocysteine $>100 \mu$ mol/L.

On the basis of genotype classification, 121 patientyears of treatment were recorded in 11 homozygotes for the I278T mutation, during which, in one patient (7), peripheral arteriosclerosis, already present at diagnosis, proceeded to development of an abdominal aortic aneurysm that required vascular surgery. During 257 patient-years of treatment in patients with another genotype, one patient (25) who suffered from a cerebrovascular event before diagnosis died, at age 42 years, of a myocardial infarction. In summary, in 378 patient-years of homocysteine-lowering treatment, only two vascular events were recorded. From the data of Mudd et al. (1985), an expected number of primary vascular events of 2 per 25 years can be calculated for untreated patients. Without treatment in our group of 27 homocystinurics, at least 30 vascular events would have been expected $(P < .01)$. During long-term followup, no first involvement of any involved organ system was noticed. Furthermore, no deterioration of clinical symptoms was recorded, except for recurrent depressive episodes in two patients (2 and 15).

Discussion

In the present study, 42 independent homocystinuric alleles were investigated, which led to the identification of 10 different mutations (table 1). The 833T \rightarrow C (I278T) mutation, first recognized by Kozich and Kraus (1992) in heterozygous state in a pyridoxine-responsive patient, was found to be the most prevalent mutation in Dutch homocystinurics: 23 (55%) of the 42 independent Dutch homocystinuric alleles carried this mutation.

Seven mutated constructs have thus far been functionally assayed in an *E. coli* expression system, which shows that all mutations tested, except D444N (Kluijtmans et al. 1996*a*), had a detrimental effect on CBS activity. The latter indicates that these mutations are likely disease-causing ones and not benign polymorphisms. The R125W and T434N mutations, detected in the two siblings of pedigree 7, still need to be analyzed in an expression system. Another mutation (R125Q) affecting the same arginine residue has been found independently by two other groups and was shown to inactivate CBS completely (Marble et al. 1994; Sebastio et al. 1995). The R369C and R491C mutations in *cis* showed a relatively high residual CBS activity of ∼10% of the control mean and are compatible with a relatively late onset of disease. Recently, R369C was postulated to be a rare polymorphism not affecting CBS function in a yeast expression system (Kim et al. 1997). The separation of both mutations by in vitro mutagenesis and expression in an *E. coli* expression system will clarify their individual effect on CBS activity.

We did not detect the $919G\rightarrow A$ mutation, which is very frequent among patients of Celtic origin with pyridoxine-nonresponsive homocystinuria (Gallagher et al. 1995). A 797G \rightarrow A mutation, a frequent cause of pyridoxine-responsive homocystinuria in Norwegian patients (Kim et al. 1997), was also not observed in these Dutch patients with homocystinuria. In a survey of 14 Italian families with predominantly pyridoxine-responsive homocystinuria patients, Sebastio et al. (1995) also did not detect the Celtic $919G \rightarrow A$ mutation and frequently observed the 833T \rightarrow C and 341C \rightarrow T transitions (in 9 and 4 of 36 alleles, respectively). Unlike $833T\rightarrow C$,

the other common CBS mutations may be of more recent genetic origin and have not spread through different populations yet. On the other hand, our observation of different haplotypes in homozygotes for $833T\rightarrow C$ mutation indicates that there is not one common ancestor haplotype in these Dutch patients (data not shown).

At diagnosis, no significant differences were observed in either biochemical or clinical expression of homocystinuria between homozygotes for I278T and patients with another genotype. Obviously, the homocystinuric phenotype is not only dependent on CBS genotype but is also influenced by other genetic and/or environmental factors. Recently, both factor V Leiden (Mandel et al. 1996) and thermolabile methylenetetrahydrofolate reductase (Kluijtmans et al. 1998) have been postulated as genetic factors modifying thromboembolic risk in homocystinurics, and, obviously, other factors may exist as well. Furthermore, although we did not observe multiple mutations in *cis* by sequencing analysis (except in patient 22), additional mutations in certain alleles might have influenced our genotype-phenotype correlation and might explain why our I278T homozygotes are more severely affected than those described by Shih et al. (1995).

In the present study, we subdivided our patients into only two groups with regard to pyridoxine responsiveness, to maintain statistical power in our analysis. On the basis of their biochemical in vivo response to large doses of pyridoxine, 48% ($n = 14$) of the Dutch homocystinurics were categorized as responsive and 31% $(n = 9)$ as nonresponsive. However, according to the classification of Brenton and Cusworth (1971), only two (7%) patients (17 and 18) of these nine would be unequivocal nonresponders; the others (24%) would be considered partially pyridoxine responsive. Not only do our observations strongly deviate from those of Mudd et al. (1985), who observed virtually equal proportions of pyridoxine-responsive and -nonresponsive patients, but they also clearly indicate the relatively mild nature of CBS deficiency in the Netherlands. The absence, in the Netherlands, of a newborn screening program for homocystinuria, by which patients with pyridoxine-nonresponsive homocystinuria, in particular, are detected (Mudd et al. 1985, 1995), might have contributed to this aberrant distribution. Nonresponders may have remained undiagnosed and lost to follow-up, because of early mortality or admittance to an institution for mentally retarded patients. The high prevalence of I278T among Dutch patients with homocystinuria might also explain the high pyridoxine responsiveness among these individuals. Seven homozygotes for this mutation showed pyridoxine responsiveness, and three individuals were classified as nonresponders. The latter finding seems to be in contradiction with the finding of pyridoxine responsiveness in three homocystinuria patients homozygous for the I278T mutation reported elsewhere (Sebastio et al. 1995; Shih et al. 1995), although in those reports no clear definitions of responsiveness were provided. The homocysteine concentrations upon pyridoxine treatment in our I278T homozygotes classified as nonresponders were only marginally above the cut-off values of 20 μ mol/L (i.e., 28 μ mol/L and 32 μ mol/L, non–protein-bound homocysteine) or 50 μ mol/L (i.e., 61 μ mol/L, total plasma homocysteine) and completely normalized upon extended treatment. So, in the present study, all homozygotes for the I278T mutation responded very successfully to homocysteine-lowering treatment in contrast to non-I278T homozygotes, of whom only 7 of 17 patients showed such beneficial biochemical response.

Homocysteine concentrations during long-term treatment showed a clear distinction between both genotype groups. Homozygotes for the I278T mutation not only had a lower mean plasma homocysteine concentration than patients with another genotype, but, on this extended treatment, the majority of this group continuously had non-protein-bound homocysteine levels <20 μ mol/L or total plasma homocysteine <50 μ mol/L. In the other genotype group, the majority of the patients persistently exceeded these limits.

Cruysberg (1996) reported that long-term therapy of Dutch homocystinurics prevented the development and progression of myopia and ectopia lentis. None of the patients without ectopia lentis at diagnosis developed this abnormality during follow-up. Furthermore, there was no progression in the degree of myopia in myopic patients. The clinical beneficial effect of the homocysteine-lowering treatment was also obvious from the overall number of vascular accidents after start of treatment. Only two vascular accidents were recorded in 378 patient-years of treatment, whereas, if untreated, at least 30 would have been expected $(P < .01)$, according to the clinical data on the natural history in homocystinuria presented elsewhere by Mudd et al. (1985). Moreover, skeletal abnormalities presented at diagnosis in high frequencies in these patients did not deteriorate during this follow-up period. Osteoporosis persisted in most cases but did not worsen, and no pathological fractures were observed (G. H. J. Boers, unpublished data). A longer follow-up in both genotype groups will be necessary to determine whether I278T homozygotes respond clinically more favorably to homocysteine-lowering treatment than do other patients with homocystinuria.

In summary, the I278T mutation is the most predominant mutation among Dutch homocystinurics. At diagnosis, no significant differences in clinical and biochemical expression were noticed between patients of both genotype groups, but homozygotes for I278T showed a higher sensitivity to homocysteine-lowering treatment by pyridoxine with or without folic acid and

betaine. The clinical efficacy of such therapy in homocystinurics emphasizes the necessity to diagnose CBS deficiency and initiate treatment as early as possible, to reduce the risk of cardiovascular disease and other sequelae of this inborn error of methionine metabolism.

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